

Original Article

Prevalence of α -Thalassemia in Malaria Patients in the Western Border of Thailand

Jiraporn Kuesap¹, Wanna Chaijaroenkul², Anurak Cheoymang¹ and Kesara Na-Bangchang²

¹Faculty of Allied Health Sciences; ²Chulabhorn International College of Medicine (Center of Excellence in Pharmacology and Molecular Biology of Malaria and Cholangiocarcinoma), Thammasat University, Pathumthani, Thailand

Abstract: Malaria remains a major public health problem in Thailand particularly in relation to infections caused by *Plasmodium falciparum* and *Plasmodium vivax*. Thalassemia is also commonly found in malaria endemic areas. In Thailand, the common α -thalassemia variants reported are α -thalassemia-1 (SEA and THAI deletion) and α -thalassemia-2 (3.7 and 4.2 deletion). This study was conducted in 2013 to investigate the prevalence of α -thalassemia in a total of 71 malaria patients residing along the Thai-Myanmar border at Mae Sot district, Tak province, Thailand. The four common α -thalassemia variants were amplified using multiplex polymerase chain reaction (PCR). Only the 3.7 kb deletion α -thalassemia-2 was found in the samples with prevalence of 19.7%. Results from this preliminary study may indirectly imply the protective effect of α -thalassemia-1 against malaria infection.

Keywords: ● Malaria ● Thalassemia ● α -thalassemia

RTA Med J 2014;67:149-54.

Introduction

Malaria is one of the parasitic diseases which remain a major public health problem in several tropical countries including Thailand. The infections caused by *Plasmodium falciparum* and *Plasmodium vivax* constitute a major burden leading to high morbidity and mortality in populations in several endemic areas. In Thailand, malaria spreads throughout the country particularly along the international borders, with the highest incidence in Tak province, in the western border of the country¹. Thalassemia is the hemoglobinopathy that is also widely

distributed in most malaria endemic areas². This genetic disease is a result of the alteration in the globin chain of hemoglobin, a tetramer molecule consisting of two each of α - and β -globin chains. The α -thalassemia leads to a decrease in the synthesis of one or both of the α -globin chains on the chromosome 16p13.3³. This α -globin gene deletion is associated with the development of a mild form α -thalassemia-2 ($-\alpha/\alpha\alpha, -\alpha/-\alpha$) or a severe form α -thalassemia-1 ($--/\alpha\alpha, --/--$). The prevalence of α -thalassemia is relatively high in several malaria endemic areas including the Mediterranean, Southeast Asia, Africa, and the Indian subcontinent⁴. Knowledge on the prevalence of α -thalassemia and its protective effect on malaria disease development, progression, and severity would be useful in providing guidance to malaria control policy. To date, the prevalence of

Received 19 November 2014 Accepted 18 December 2014

Requests for reprints should be addressed to Prof. Dr. Kesara Na-Bangchang, Chulabhorn International College of Medicine, Thammasat University (Rangsit Campus), Klongluang, Pathumthani, 12121, Thailand.
E-mail: kesaratmu@yahoo.com,

α -thalassemia including its association with malaria infection in Thailand has not been reported. The aim of this preliminary study was to investigate the prevalence of α -thalassemia in malaria patients in Mae Sot district, Tak province on the western border of Thailand.

Materials and Methods

Study subjects and sample collection

A total of 71 blood samples were collected from patients (4 Thais and 67 Burmese) infected with *P. falciparum* (n = 26), *P. vivax* (n = 43), and mixed infection of both species (n = 2) who attended malaria clinics in Mae Sot district, Tak province, Thailand in 2013. Approval of the study protocol was obtained from the Ethics Committee of Thammasat University. Blood samples were stained with Giemsa and microscopically examined for the presence of malaria parasites.

Extraction of parasite genomic DNA

Parasite genomic DNA was extracted from whole blood using QIAamp DNA extraction mini-kit (QIAGEN, CA, USA) according to standard protocol and used as a template for polymerase chain reaction (PCR) amplification.

Amplification of α -thalassemia

Mutations of six variants of the genes encoded α -thalassemia were amplified using three sets of multiplex PCR⁵⁻⁷. These included SEA and THAI variants of α -thalassemia-1, the 3.7 and 4.2 kb deletion variants of α -thalassemia-2, Hb Constant Spring (HbCS), and Hb Pakse (HbPS).

Identification of α -thalassemia-1 with SEA (Southeast Asia) and THAI deletions was performed using T1 (5'-TGACTGCATCATAATTCAGCAG-3') and T2 (5'-TGAGGCAGGAGATTGCTTGA-3') primers. The three primers A7 (5'-CTCTGTGTTCTC AGTATTGGAG-3'), A9 (5'-ATATATGGGTCTGGAAGTGTATC-3'), and A1B (5'-GG TTCCCTGAGCCCC GACACG-3') were used to amplify the SEA deletion (660 bp) and internal control (314 bp)^{5,6}.

The PCR reaction mixture contained 50-100 ng DNA, 75 mM Tris-HCl (pH 8.0), 2 mM MgCl₂, 50 mM KCl, 20 μ M (NH₄)₂SO₄, 200 mM of each dNTP, 67.5 pmol of A7 primer, 33.8 pmol of A9 primer, 2 pmol of A1B primer, 6 pmol of T1 and T2 primers, 1 M betaine, 1% DMSO, and 2.5 units Taq polymerase (Promega Co., Madison, USA). The PCR condition consisted of 1 cycle of 94°C for 3 min, 35 cycles of 94°C for 40 s, 61°C for 20 s, 72°C for 2 min, and final extension at 72°C for 2 min. The products were separated on a 2% agarose gel containing ethidium bromide.

The identification of α -thalassemia-2 with 3.7 and 4.2 kb deletions was performed using three sets of primers, i.e., α G26 (5'-CTCAACAGATGCCAGCCAAA CAGAC-3')/ α G27 (5'-GAACCATTTACACCAAGTGAA GG ACG-3'), A (5'-CCCAGAGCCAGGTTTGTATTATC TGT-3')/B (5'-GAGGCCCAAGGGGCAAGAAGCAT-3'), and C (5'-GCTAGAGCATTTGGTGGTCATGCC-3')/D (5'-TTCTGACTCTGCCCCACAGCCTGA-3'), to amplify the internal control (933 bp), 3.7 kb deletion (1,779 bp), and 4.2 kb deletion (1,529 bp), respectively⁷. The PCR reaction mixture contained 0.1 μ g DNA, 30 pmol each of the A, B, C, and D primers, 15 pmol each of the α G26 and α G27 primers, 200 μ M dNTPs, 75 mM Tris-HCl (pH 8.0), 50 mM KCl, 2 mM MgCl₂, 1 M betaine, 5% dimethylsulfoxide, and 2 units of Taq DNA polymerase (New England Biolabs Inc., MA, USA). The PCR condition consisted of 1 cycle of 94°C for 3 min, 10 cycles of 94°C for 30 s, 60°C for 30 s, 72°C for 2 min, and 20 cycles with 20 s increments in the extension step of each cycle. The products were separated on a 1.5% agarose gel containing ethidium bromide.

The identification of HbCS and HbPS mutations was performed using α G2 (5'-GCTGACCTCCAAATACCGTC-3')/C3 (5'-CCATTGTTGGCACATTCCGG-3'), and α G17 (5'-AGATGGCGCCTTCCTCTCAGG-3')/ α G18 (5'-ACGGCT ACCGAGGCTCCAGCA-3'), to amplify α^{CS} and α^{PS} .

mutations, respectively⁸. The PCR reaction mixture contained 0.1 μ g DNA, 37.5 pmol of α G2 primer, 30 pmol of C3 primer, 3.75 pmol of α G17 primer, 30 pmol α G18 primer, 200 μ M dNTPs, and 2 units of Taq DNA polymerase (Promega Co., Madison, USA) in 10 mM Tris-HCl, 50 mM KCl, 0.1% Triton X-100, 3.0 mM $MgCl_2$, 1 M betaine, and 4.8% dimethylsulfoxide. The PCR condition consisted of 1 cycle of 94°C for 30 min, 30 cycles of 94°C for 1 min, and 65°C for 1 min. The products were separated on a 1.5% agarose gel containing ethidium bromide.

Statistical analysis

Statistical analysis was performed using the SPSS statistical package (version 12.0 SPSS Inc., IL, USA). Difference of qualitative and quantitative data between groups was analyzed using chi-square test and ANOVA with Tukey analysis, respectively. Statistical significance level was set at $\alpha = 0.05$.

Results

The prevalence of α -thalassemia in 71 patients with malaria infection in Mae Sot district, Tak province, Thailand was 19.7%. Four types of gene mutations were observed, i.e., homozygous α -thalassemia-2, homozygous α -thalassemia-2 with Hb E, heterozygous α -thalassemia-2, and heterozygous α -thalassemia-2 with β -thalassemia (Table 1). The major types were heterozygous α -thalassemia-2 with and without β -thalassemia (16.9%). All patients with α -thalassemia-2 had a 3.7 kb gene deletion. The α -thalassemia-1 was not detected in any sample. The hematological parameters including hemoglobin, hematocrit, mean corpuscular volume (MCV), and mean corpuscular hemoglobin (MCH) are presented in Table 1. There was no significant association between α -thalassemia types and malaria species, patient gender, hemoglobin, and hematocrit values. The median values of MCV and MCH of patients with heterozygous α -thalassemia-2

Table 1 Prevalence of α -thalassemia 2, malaria species infected, patient gender, and hematological parameters in 71 malaria patients. The hematological parameters are presented as median (minimum-maximum) values.

| Genotype of thalassemia | Number (%) | Malaria Type | | | Gender | | Hb (g/dL) | Hct (%) | MCV (fl) | MCH (pg) |
|---|------------|--------------|----|-----|--------|--------|------------------|------------------|-------------------------------|-------------------------------|
| | | PF | PV | MIX | Male | Female | | | | |
| Homozygous α -thal2 | 1 (1.4) | 1 | 0 | 0 | 1 | 0 | 12.0 (12.0-12.0) | 34.3 (34.3-34.3) | 65.0 (65.0-65.0) | 22.9 (22.9-22.9) |
| Homozygous α -thal2 with HbE | 1 (1.4) | 0 | 1 | 0 | 1 | 0 | 7.6 (7.6-7.6) | 24.0 (24.0-24.0) | 62.0 (62.0-62.0) ^c | 19.7 (19.7-19.7) |
| Heterozygous α -thal2 | 9 (12.7) | 2 | 7 | 0 | 5 | 4 | 12.0 (5.1-14.3) | 36.5 (15.9-44.1) | 74.0 (53.0-91.0) | 24.1 (15.2-30.2) |
| Heterozygous α -thal2 with β -thal | 3 (4.2) | 1 | 2 | 0 | 1 | 2 | 10.9 (9.8-11.1) | 33.6 (28.3-34.3) | 67.0 (60.0-67.0) ^a | 21.3 (19.6-22.7) ^b |
| Malaria without α -thalassemia | 57 (80.3) | 22 | 33 | 2 | 39 | 18 | 12.2 (3.4-16.4) | 37.1 (10.7-49.1) | 80.0 (50.0-101.0) | 26.8 (18.7-32.3) |

PF = *P. falciparum*; PV = *P. vivax*; Hb = hemoglobin; Hct = hematocrit; thal = thalassemia

^a Statistically significant difference from patients without thalassemia ($p = 0.005$)

^b Statistically significant difference from patients without thalassemia ($p = 0.001$)

^c Statistically significant difference from patients without thalassemia ($p = 0.013$)

combined with β -thalassemia were significantly lower than those without thalassemia (MCV $p = 0.005$; MCH $p = 0.001$). Furthermore, the median MCV of patients with homozygous α -thalassemia-2 combined with Hb E was significantly lower than patients without thalassemia ($p = 0.013$).

Discussion and Conclusion

The α -Thalassemia is commonly found in Southeast Asia and China, and the most common variants of which are α -thalassemia-1-^{SEA} and α -thalassemia-2- α ^{3,7}. In Thailand, the prevalence of α -thalassemia in different regions of the country varies from 2.5 to 30%^{9,10}. Results of the present study revealed only the 3.7 kb deletion α -thalassemia-2 in the western region of the country at frequency of 19.7%. The prevalence of α -thalassemia in the population in the western region of Thailand has not been previously reported. This frequency is similar to what was reported in the population in the northeastern region of the country (17.5%)¹¹. As most of the study population was Burmese, this observed prevalence could have been confounded by influence of ethnicity. An average prevalence of 5-37.5% of α -thalassemia has been reported in Burmese population^{12,13}. The most common α -thalassemia variant (29.7%) observed in the Shan State close to the northwest border of Thailand is heterozygous α -thalassemia-2¹³.

The α -thalassemic red cells were reported to protect the malaria infected host against disease development and progression^{14,15}. This protective effect was supported by the reports from Tanzania and Kenya^{16,17}, Vanuatu¹⁸, and Ghana¹⁹. The proposed mechanism is the reduction of the ability of the infected thalassemic red cells to form rosettes with the uninfected red cells and thus preventing the sequestration of malaria parasites in blood capillaries²⁰. In addition, thalassemic red cells increase the amounts of immunoglobulin, immune recognition,

resulting in the acceleration of the clearance of infected red cells²¹. The α -thalassemic red cells also promote the pro-inflammatory effect of cytoadherence²². The α -Thalassemia-2 does not appear to protect malarial infection including progression to symptomatic or high parasitemia, but reduce the risk of developing severe malaria¹⁵. The study in Melanesian children showed the lack of protective effect of α -thalassemia-2 against uncomplicated *P. falciparum*²³. The observation of only α -thalassemia-2 in this patient population may indirectly imply the protective effect of α -thalassemia-1 against malaria infection. However, the limitation of the current study is the lack of non-malaria patient population, to investigate the relationship between malaria and thalassemia. Further study with increasing sample size is needed to confirm the protective effect of α -thalassemia on malaria.

Acknowledgements

The study was supported by Thammasat University (TU) Research Fund under the TU Research Scholar Contract No. 57/2555. KN was supported by the Commission on Higher Education (National University Project), Ministry of Education, and Thammasat University (Center of Excellence in Pharmacology and Molecular Biology of Malaria and Cholangiocarcinoma).

References

1. Na-Bangchang K, Congpuong K. Current malaria status and distribution of drug resistance in East and Southeast Asia with special focus to Thailand. *Tohoku J Exp Med* 2007;211:99-113.
2. Vento S, Cainelli F, Cesario F. Infections and thalassaemia. *The Lancet Infectious Diseases* 2006;6:226-33.
3. Yuthavong Y, Wilairat P. Protection against malaria by thalassaemia and haemoglobin variants. *Parasitology today* 1993;9:241-5.
4. Weatherall DJ, Clegg JB. *The Thalassaemia Syndromes*, Chapter 1, Blackwell Science, Oxford, UK, 4th edition, 2001.
5. Eng B, Patterson M, Borys S, Chui DH, Wayne JS. PCR base diagnosis of the Filipino (- - FIL) and Thai (- - THAI) α -thalassemia-1 deletions. *Am J Hematol* 2000;63:54-6.

6. Panyasai S, Sringam P, Fucharoen G, Sanchaisuriya K, Fucharoen S. A simplified screening for α -thalassemia 1 (SEA type) using a combination of a modified osmotic fragility test and a direct PCR on whole blood cell lysates. *Acta Haematol* 2002;108:74-8.
7. Chaibunruang A, Kampean R, Fucharoen G, Fucharoen S. Genetic heterogeneity of hemoglobin AEBart's disease: a large cohort data from a single referral center in northeast Thailand. *Blood Cells Mol Dis* 2014;52:176-80.
8. Fucharoen S, Sanchaisuriya K, Fucharoen G, Panyasai S, Devenish R, Luy L. Interaction of hemoglobin E and several forms of alpha-thalassemia in Cambodian families. *Haematologica* 2003;88:1092-8.
9. Fucharoen S, Winichagoon P. Hemoglobinopathies in Southeast Asia. *Hemoglobin* 1987;11(1):65-88.
10. Fucharoen S. Thalassemia and hemoglobinopathies in northeast Thailand and greater Mekong sub-region. *Thai J Genet* 2013;S(1):17-22.
11. Tritipsombut J, Sanchaisuriya K, Phollarp P, et al. Micromapping of thalassemia and hemoglobinopathies in different regions of northeast Thailand and Vientiane, Laos People's Democratic Republic. *Hemoglobin* 2012;36:47-56.
12. Weatherall DJ. Thalassemia in the next millennium: keynote address. *Ann N Y Acad Sci* 1998;850:1-9.
13. Than AM, Harano T, Harano K, Myint AA, Ogino T, Okada S. High incidence of 3-thalassemia, hemoglobin E, and glucose-6-phosphate dehydrogenase deficiency in populations of malaria-endemic southern Shan State, Myanmar. *Int J Hematol*. 2005;82:119-23.
14. Williams TN, Mwangi TW, Wambua S, et al. Negative epistasis between the malaria-protective effects of alpha+-thalassemia and the sickle cell trait. *Nature genetics* 2005a;37:1253-7.
15. Wambua S, Mwangi TW, Kortok M, et al. The effect of alpha+-thalassaemia on the incidence of malaria and other diseases in children living on the coast of Kenya. *PLoS medicine* 2006;3:e158.
16. Veenemans J, Andang'o PE, Mbugi EV, et al. Alpha+ -thalassemia protects against anemia associated with asymptomatic malaria: evidence from community-based surveys in Tanzania and Kenya. *J Infect Dis* 2008;198:401-8.
17. Williams TN, Wambua S, Uyoga S, et al. Both heterozygous and homozygous alpha+ thalassemias protect against severe and fatal *Plasmodium falciparum* malaria on the coast of Kenya. *Blood* 2005;106:368-71.
18. Ubalee R, Tsukahara T, Kikuchi M, et al. Associations between frequencies of a susceptible TNF-alpha promoter allele and protective alpha-thalassaemias and malaria parasite incidence in Vanuatu. *Trop Med Int Health* 2005;10:544-9.
19. Opoku-Okrach C, Gordge M, Kweku Nakua E, Agbenyega T, Parry M, Robertson C, Smith CL. An investigation of the protective effect of alpha+-thalassaemia against severe *Plasmodium falciparum* amongst children in Kumasi, Ghana. *Int J Lab Hematol* 2013, doi:10.1111/ijlh.12122.
20. Carlson J, Nash GB, Gabutti V, al-Yaman F, Wahlgren M. Natural protection against severe *Plasmodium falciparum* malaria due to impaired rosette formation. *Blood* 1994;84:3909-14.
21. Luzzi GA, Merry AH, Newbold CI, et al. Surface antigen expression on *Plasmodium falciparum* -infected erythrocytes is modified in alpha- and beta- thalassemia. *J Exp Med* 1991;173:785-91.
22. Krause MA, Diakite SA, Lopera-Mesa TM, et al. α -thalassemia impairs the cytoadherence of *Plasmodium falciparum*-infected erythrocytes. *PLoS One* 2012;7:e37214.
23. Rosanas-Urgell A, Senn N, Rarau P, et al. Lack of associations of α (+)-thalassemia with the risk of *Plasmodium falciparum* and *Plasmodium vivax* infection and disease in a cohort of children aged 3-21 months from Papua New Guinea. *Int J Parasitol* 2012;42:1107-13.

ความชุกของแอลฟาธาลัสซีเมีย ในผู้ป่วยมาลาเรียภาคตะวันตกของประเทศไทย

จิราภรณ์ คัญทรัพย์¹ วรณา ชัยเจริญกุล² อนุรักษ์ เชื้อมั่ง¹ และ เกศรา ณ บางช้าง²

¹คณะสหเวชศาสตร์ ²วิทยาลัยแพทยศาสตร์นานาชาติจุฬาภรณ์ มหาวิทยาลัยธรรมศาสตร์

บทคัดย่อ มาลาเรียปัญหายังคงเป็นมหาโรคสำคัญในประเทศไทยโดยเฉพาะมาลาเรียชนิดพลาสโมเดียมและไวแวกซ์ นอกจากนี้ในพื้นที่ระบาดของมาลาเรียมักพบอุบัติการณ์ของธาลัสซีเมียด้วย โดยแอลฟาธาลัสซีเมียชนิดที่พบบ่อยคือแอลฟาธาลัสซีเมีย-1 และ -2 การศึกษานี้ได้ดำเนินการในปี พ.ศ. 2556 โดยมีวัตถุประสงค์เพื่อศึกษาความชุกของแอลฟาธาลัสซีเมียในผู้ป่วยมาลาเรียที่อาศัยในพื้นที่ อำเภอมะนัง จังหวัดตาก จำนวน 71 ราย โดยศึกษาแอลฟาธาลัสซีเมียชนิดที่พบบ่อยคือ แอลฟาธาลัสซีเมีย-1 (SEA และ THAI deletion) และแอลฟาธาลัสซีเมีย-2 (3.7 และ 4.2 deletion) ด้วยวิธี multiplex polymerase chain reaction (PCR) ในประชากรที่ศึกษาพบเฉพาะแอลฟาธาลัสซีเมีย-2 ชนิด 3.7 deletion เพียงชนิดเดียว ผลการศึกษาในเบื้องต้นนี้อาจแสดงให้เห็นถึงผลในการป้องกันการติดเชื้อมาลาเรียทั้งชนิดพลาสโมเดียมและชนิดไวแวกซ์ของแอลฟาธาลัสซีเมีย-1 ในผู้ป่วยมาลาเรียที่อาศัยในพื้นที่

Keywords: ● มาลาเรีย ● ธาลัสซีเมีย ● แอลฟาธาลัสซีเมีย

เวชสารแพทย์ทหารบก 2557;67:149-54.